

Visible light induced Barton decarboxylation free of radical initiators

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Abstract: Herein, a novel protocol for visible light induced decarboxylation was reported. The method avoids the addition of radical initiators. A series of primary, secondary, and tertiary redox-active esters underwent decarboxylation efficiently, as well as amino acid and peptide derivatives. Many natural products and drug molecules containing carboxyl groups can also be transformed well. The method has mild reaction conditions, wide substrate scope, satisfactory functional group compatibility with good to excellent yields. The preliminary mechanism studies suggested that tris(trimethylsilyl)silane((TMS)₃SiH) acts not only as a hydrogen source, but also as an electron donor to form electron donor-acceptor (EDA) complex with redox-active esters, and the reaction involves the radical process.

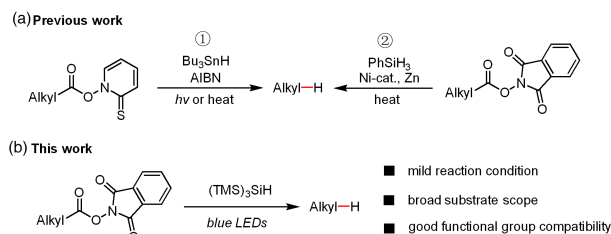
Keywords: visible light induced; electron donor-acceptor complex; decarboxylation; radical; tris (trimethylsilyl)silane

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1 Introduction

The unit of carboxylic acid is an important class of organic synthons, and commonly found in natural products and drug molecules^[1-3]. Due to its intrinsic properties of being readily available, inexpensive, bench-stable, non-toxic and easy to store, they have been widely used in organic synthetic chemistry. The named reaction of Barton decarboxylation is an effective protocol for the decarboxylation of alkyl carboxylic acids into alkanes, which involves the radical process^[4-7]. Barton esters (thiohydroxamic esters) can liberate carbon dioxide under light irradiation or heat conditions in the presence of tributyltin hydride and 2, 2'-azobis (2-methylpropionitrile) (AIBN) as the radical initiator, deliver a variety of alkanes (Scheme 1, (a) ①). Considering the toxicity of the reductant, other alternatives such as tris (trimethylsilyl) silane ((TMS)₃SiH)^[8] or chloroform^[9] have been introduced for Barton decarboxylation. However, these methods still inevitably use the low stability thiohydroxamic esters which are photo- and thermal sensitive, and the radical initiators. Therefore, it is still highly desirable to develop a much greener and more efficient method for Barton decarboxylation. For this, Okada and Oda's group developed a novel *N*-hydroxyphthalimide redox-active ester, which can be easily obtained from alkyl carboxylic acids and *N*-hydroxyphthalimide^[10]. Using 1,

6-bis (dimethylamino) pyrene (BDMAP) as the photosensitizer and *tert*-butyl mercaptan as the hydrogen source, a series of alkyl carboxylic acid derivatives can proceed decarboxylation smoothly under the irradiation of 400 W Hg lamp. Recently, Baran et al. reported a nickel-catalyzed decarboxylative protocol using PhSiH₃ as a hydrogen atom source. Various functionalized *N*-hydroxyphthalimide redox-active esters successfully delivered the reductive decarboxylation products (Scheme 1,(a) ②)^[11].



Scheme 1 Barton decarboxylation.

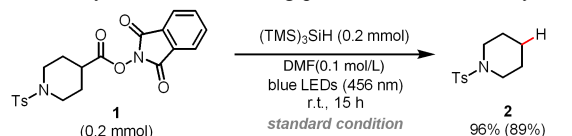
In recent years, the strategy of visible-light catalysis has received much attention in organic synthetic area, due to its ability to prompt chemists to discover new reactivity of traditional catalysts, and realize many important transformations that cannot be achieved by traditional methods^[12-17]. Our laboratory made progress in this area^[18,19], especially for visible light-induced general catalytic radical generation from electron donor-acceptor complex (EDA complex)^[20-22]. The single electron donor or electron acceptor substrate

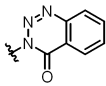

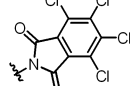
has no obvious absorption in visible light range but the resulting EDA complex does, and subsequently induces electron transfer to achieve free radical type conversion under visible light irradiation^[23,24]. Based on continuous research in this field, we found that associating the redox-active esters of *N*-acyloxyphthalimide as the electron acceptor with $(\text{TMS})_3\text{SiH}$ ^[25] as the electron donor can form the EDA complex, which can successfully undergo Barton decarboxylation process under the irradiation of 456 nm blue light-emitting diodes. The method has mild reaction conditions, wide substrate scope, good functional group compatibility. Primary, secondary, tertiary alkyl carboxylic acids, as well as amino acid and peptide derivatives smoothly underwent decarboxylation with good to excellent yields. The simple operation and high yields of this practical protocol will be a welcome advance for Barton decarboxylation.

2 Results and discussion

2.1 Screening of reaction conditions

Tab. 1 Key reaction-controlling parameters of decarboxylation.



different solvents instead of DMF							
DMA	MeCN	THF	PhCF ₃	CH ₂ Cl ₂	EtOAc	dioxane	acetone
90	50	trace	trace	trace	trace	56	52
different silanes instead of $(\text{TMS})_3\text{SiH}$							
PhSiH ₃	Ph ₂ SiH ₂	Et ₂ SiH ₂	Ph ₃ SiH	Et ₃ SiH	(EtO) ₃ SiH		
0	0	0	0	0	0		
different wavelength of visible light							
467 nm	440 nm	427 nm	390 nm				
40	95	96	93				
control experiments							
w/o light, 60 °C	$(\text{TMS})_3\text{SiH}$ (0.1 mmol)	$(\text{TMS})_3\text{SiH}$ (0 mmol)					
0	44	0					
other redox-activation groups instead of -NPhth							
							
trace	55	94 ^a					

Reaction conditions: **1** (0.2 mmol), $(\text{TMS})_3\text{SiH}$ (0.2 mmol), DMF (2.0 mL), room temperature (r.t.), 15 h, blue LEDs (456 nm). The yield was determined by ¹H NMR using diphenylmethane as internal standard. Isolated yield in parentheses.^a Reaction time: 15 min.

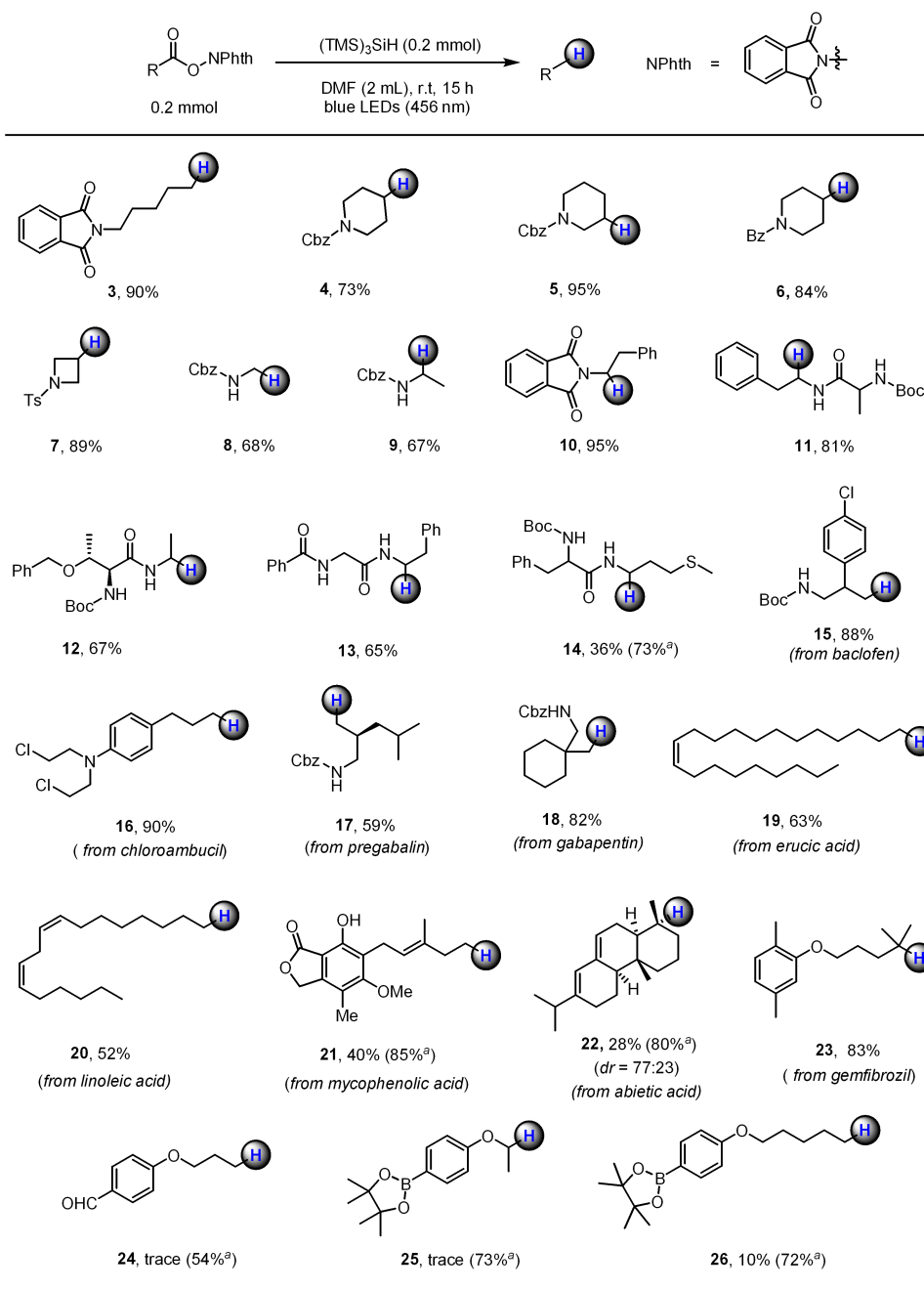
Our investigation into the key reaction-controlling parameters of decarboxylation is shown in Tab. 1. At the beginning, **1** was chosen as the model substrate. After careful optimization of all reaction parameters, the

desired product **2** was obtained in 96% yield using $(\text{TMS})_3\text{SiH}$ (0.2 mmol) as hydrogen source in DMF (*N,N*-dimethylformamide) under 456 nm blue LEDs irradiation at room temperature. Solvation has a significant influence on the assembly of electron donor-acceptor complex and an effect on the reaction outcome (row 1). When using DMA (*N,N*-dimethylacetamide) as the solvent, 90% yield of the product was obtained. When using CH₃CN, dioxane or acetone instead of DMF, the reaction proceeded in lower yields. The reaction was totally suppressed when using THF (tetrahydrofuran), PhCF₃, CH₂Cl₂ or EtOAc as the solvent. To our surprise, other silanes, such as PhSiH₃, Ph₂SiH₂, Et₂SiH₂, Ph₃SiH, Et₃SiH and (EtO)₃SiH failed to afford the desired product with the raw materials remaining intact (row 2), which indicated that $(\text{TMS})_3\text{SiH}$ acts not only as a hydrogen source, but also as the electron donor to form electron donor-acceptor (EDA) complex with **1**. Next, the wavelength of the irradiation source was investigated (row 3). The reaction performed smoothly under the wavelength of 390 nm to 456 nm, whereas the efficiency dramatically dropped when the wavelength was increased to 467 nm, only 40% yield of **2** was obtained. Control experiments showed that both irradiation and $(\text{TMS})_3\text{SiH}$ were essential for the success of this new Barton decarboxylation protocol. The yield of the desired product **2** was decreased to 44% when the amount of $(\text{TMS})_3\text{SiH}$ was reduced to 0.5 equivalent. In addition, we also investigated other types redox-active esters in this protocol (row 4). Using 3-hydroxy-1, 2, 3-benzotriazin-4 (3*H*)-one as the active group was ineffective, but 1-hydroxybenzo-triazole gave **2** in 55% yield. It was worth noting that when using tetrachloro-*N*-hydroxyphthalimide-derived redox-active esters, the desired product could be obtained equivalently within 15 min. Considering the issue of atom economy, we still chose *N*-hydroxyphthalimide types of redox-active esters in the following investigation.

2.2 Scope of the decarboxylation

With the optimized conditions in hand, we next explored the reaction scope for this hydrodecarboxylation. As shown in Tab. 2, primary, secondary and third redox-active esters of *N*-acyloxyphthalimide were readily converted to the corresponding alkanes in good to excellent yields. This protocol has good functional group tolerance, such as phthalimide (**3**), benzyloxycarbonyl (**4**, **5**), benzoyl (**6**) and *p*-toluenesulfonyl (**7**), all well-tolerated. Both six-membered ring and four-membered ring carboxylic acid-derivatives transformed smoothly (**4-7**). In addition, this protocol was also successfully applied to amino acid and peptide derivatives (**8-13**). Redox-active esters of amino acids or dipeptides with different protecting groups, such

Tab. 2 Scope of substrates



Reaction conditions: redox-active esters (0.2 mmol, 1.0 equiv.), $(\text{TMS})_3\text{SiH}$ (0.2 mmol), DMF (2 mL), 15 h, r.t., blue LEDs (456 nm). Isolated yield. ^a NaI (10 mol%), PPh_3 (10 mol%).

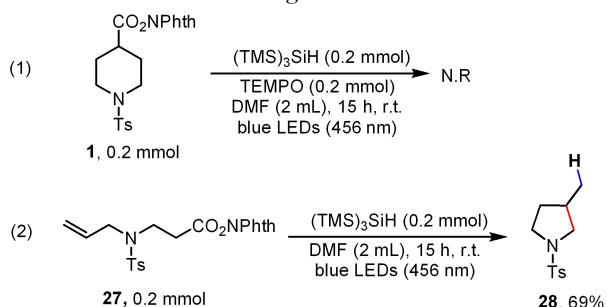
as benzyloxycarbonyl, phthalimide, and tert-butoxycarbonyl group, were amenable substrates, delivered 65% ~ 95% isolated yield of the desired product. Decarboxylative hydrogenation product **14** was obtained only in 36% isolated yield using this method. To our delight, we found the yield can be increased to 73% by adding a catalytic amount of sodium iodide and triphenylphosphine^[18].

To further test the practicability of this method, we investigated a series of natural products and drug

molecules containing carboxylic acid groups (**15-23**). The drug molecules derivatives, such as baclofen, chloroambucil, pregabalin, gabapentin, mycophenolic acid and gemfibrozil also underwent decarboxylation smoothly to the corresponding alkanes. Additionally, various carbonyl functional groups, including aryl chloride (**15**), alkyl chloride (**16**), *N,N*-dialkyl arylamine (**16**), ester (**21**) and aryl ether (**23**), were all tolerated under this condition. It was worth noting that the unprotected acidic phenolic hydroxyl group in

mycophenolic acid-derived Barton ester (**21**) also remained intact, and the yield could be successfully increased to 85% by adding a catalytic amount of sodium iodide and triphenylphosphine. Natural products, such as erucic acid, linoleic acid and abietic acid could be transformed smoothly, and the alkene group susceptible to free radicals also remained intact. All these results highlight the high chemoselectivity and functional group tolerance of the method. It should be noted that the activity of the reaction was totally suppressed when using the substrates containing aromatic aldehydes (**24**) and aromatic boronic esters (**25**, **26**). It is possible that these functional groups affect the formation of EDA complexes. Gratifyingly, by adding a catalytic amount of sodium iodide and triphenylphosphine, these types of substrates could smoothly undergo decarboxylation.

2.3 Mechanistic investigation



To further shed light on the mechanism of this method, the radical inhibitor 2, 2, 6, 6-tetramethylpiperidinyloxy (TEMPO) was added to the model reaction system (Eq. (1)), and the formation of product **2** was totally suppressed. Furthermore, the radical-clock experiment was conducted using **27** as the substrate, giving ring-closing pyrrolidine product **28** as exclusive product. These results all support the involvement of a free radical mechanism in this reaction^[26].

A large number of literatures have been reported about the reaction with $(\text{TMS})_3\text{SiH}$ participating as a radical-based reducing agent, with heating condition or

with additional radical initiators (e. g. AIBN)^[25]. However, the radical initiator was not necessary in this method. We speculated that $(\text{TMS})_3\text{SiH}$ could act as the electron donor to form the chromophore (EDA complex) with *N*-acyloxypthalimide redox-active ester as the electron acceptor through weak interaction. Under visible light irradiation, intermolecular electron transfer could occur between $(\text{TMS})_3\text{SiH}$ and *N*-acyloxypthalimide redox-active ester, subsequently generating alkyl radicals with liberating carbon dioxide, and then reacting with the reducing agent $(\text{TMS})_3\text{SiH}$ to form the decarboxylated product.

In order to further verify the above speculation, the ultraviolet-visible spectrophotometric experiment was carried out under the condition of maintaining the original concentration of the model reaction (Fig. 1). The experimental results showed that $(\text{TMS})_3\text{SiH}$ has almost no absorption above the wavelength of 400 nm. However, an obvious redshift absorption of the mixed solution of $(\text{TMS})_3\text{SiH}$ with compound **1** was observed compared to pure **1** solution, which confirmed that the chromophore was formed between $(\text{TMS})_3\text{SiH}$ and **1** in DMF.

3 Conclusions

In summary, an efficient visible light induced Barton decarboxylation has been developed without using a radical initiator. The reaction proceeds under mild conditions with excellent functional group compatibility and broad substrate scope. Primary, secondary, and tertiary alkyl acid-derivatives underwent decarboxylation efficiently, as well as amino acid and peptide derivatives. Preliminary mechanism studies suggested that $(\text{TMS})_3\text{SiH}$ acts not only as a radical-based reducing agent, but also as the electron donor to form electron donor-acceptor (EDA) complex with redox-active esters.

4 Experimental section

4.1 General procedure

General procedure A: Redox active ester (1.0 eq., 0.2 mmol) was placed in a 10 mL transparent Schlenk tube equipped with a stirring bar. The tube was evacuated and filled with argon (three times). To these solids, anhydrous DMF (2.0 mL) and $(\text{TMS})_3\text{SiH}$ (1.0 eq., 0.2 mmol) were added via a gastight syringe under argon atmosphere. The reaction mixture was stirred under irradiation with blue LEDs (456 nm, distance app. 3.0 cm from the bulb), maintained at approximately room temperature in the air-conditioned room of 25°C. After 15 h, the mixture was quenched with 5 mL water, then extracted with ethyl acetate (3 x 10 mL). The organic layers were combined and concentrated under vacuo. The product was purified via flash column

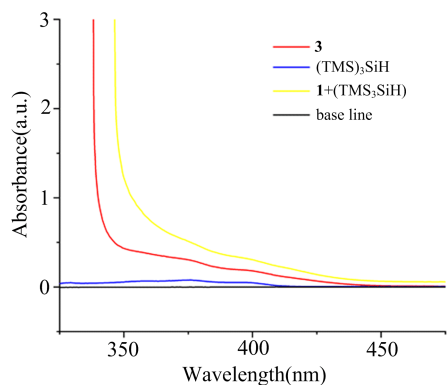


Fig. 1 UV-vis spectrum.

chromatography on silica gel (petroleum ether/ethyl acetate=50:1~2:1).

General procedure B: Redox active ester (1.0 eq., 0.2 mmol), NaI (20 mol%), PPh₃ (20 mol%) were placed in a 10 mL transparent Schlenk tube equipped with a stirring bar. The tube was evacuated and filled with argon (three times). To these solids, anhydrous DMF (2.0 mL) and (TMS)₃SiH (1.0 eq., 0.2 mmol) were added via a gastight syringe under argon atmosphere. The reaction mixture was stirred under irradiation with blue LEDs (456 nm, distance app. 3.0 cm from the bulb), maintained at approximately room temperature in the air-conditioned room of 25 °C. After 15 h, the mixture was quenched with 5 mL water, then extracted with ethyl acetate (3 x 10 mL). The organic layers were combined and concentrated under vacuo. The product was purified via flash column chromatography on silica gel (petroleum ether/ethyl acetate=50:1~2:1).

4.2 Characterization data for products

1-tosylpiperidine (2)^[11]: obtained following the procedure A, 95% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.64 (d, *J*=8.2 Hz, 2H), 7.32 (d, *J*=8.1 Hz, 2H), 3.01–2.92 (m, 4H), 2.43 (s, 3H), 1.64 (dt, *J*=11.3, 5.8 Hz, 4H), 1.46–1.35 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 143.3, 133.2, 129.6, 127.7, 46.9, 25.2, 23.5, 21.5.

2-pentylisoindoline-1, 3-dione (3)^[27]: obtained following the procedure A, 45.4 mg, 90% yield, white solid. The compound data was in agreement with the literature (Ref: *J. Am. Chem. Soc.* 2016, 138, 696–702). ¹H NMR (400 MHz, CDCl₃) δ 7.69–7.60 (m, 2H), 7.56–7.49 (m, 2H), 3.51–3.45 (m, 2H), 1.58–1.37 (m, 2H), 1.23–1.04 (m, 4H), 0.70 (t, *J*=6.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 168.5, 133.8, 132.2, 123.1, 38.0, 29.0, 28.3, 22.3, 13.9.

benzyl piperidine-1-carboxylate (4)^[6]: obtained following the procedure A, 32.0 mg, 73% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.72–7.00 (m, 5H), 5.12 (s, 2H), 3.63–3.19 (m, 4H), 1.78–1.42 (m, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 155.3, 137.0, 128.4, 127.9, 127.8, 66.9, 44.9, 25.7, 24.4.

phenyl (piperidin-1-yl) methanone (6)^[28]: obtained following the procedure A, 31.8 mg, 83% yield, colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ 7.42–7.36 (m, 5H), 3.71 (s, 2H), 3.34 (s, 2H), 1.75–1.46 (m, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 170.4, 136.4, 129.4, 128.4, 126.8, 48.8, 43.1, 26.4, 25.9, 24.6.

1-tosylazetidine (7)^[29]: obtained following the procedure A, 37.6 mg, 89% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.73 (d, *J*=8.3 Hz, 2H), 7.38 (d, *J*=8.4 Hz, 2H), 3.87–3.71 (m, 4H), 2.47 (s, 3H), 2.18–1.94 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 144.0, 131.5, 129.7, 128.4, 50.9, 21.6, 15.3.

benzyl methylcarbamate (8)^[30]: obtained following the

procedure A, 22.5 mg, 68% yield, colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ 7.61–7.26 (m, 5H), 5.10 (s, 2H), 4.76 (br s, 1H), 2.79 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 157.0, 136.6, 128.5, 128.1, 66.7, 27.6.

benzyl ethylcarbamate (9)^[31]: obtained following the procedure A, 24.0 mg, 67% yield, colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ 7.49–7.28 (m, 5H), 5.09 (s, 2H), 4.78 (br, 1H), 3.23 (br, 2H), 1.13 (t, *J*=7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 156.3, 136.7, 128.5, 128.1, 128.1, 66.6, 35.9, 15.3.

2-phenethyl-1H-indene-1,3(2H)-dione (10): obtained following the procedure A, 44.7 mg, 95% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.82 (m, 2H), 7.74–7.67 (m, 2H), 7.33–7.17 (m, 5H), 3.92 (m, 2H), 3.04–2.84 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 168.2, 138.0, 133.9, 132.1, 128.9, 128.6, 126.7, 123.2, 39.3, 34.6. HRMS (ESI) calcd. for C₁₆H₁₃NO₂Na⁺: [M+Na]⁺: 274.0844, found: 274.0832

tert-butyl (1-oxo-1-(phenethylamino) propan-2-yl) carbamate (11)^[32]: obtained following the procedure A, 47.4 mg, 81% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.34–7.15 (m, 5H), 6.33 (br s, 1H), 5.08 (br s, 1H), 4.11 (s, 1H), 3.62–3.41 (m, 2H), 2.81 (t, *J*=7.1 Hz, 2H), 1.43 (s, 9H), 1.31 (d, *J*=7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 172.7, 155.5, 138.7, 128.7, 128.6, 126.5, 80.0, 50.1, 40.7, 35.7, 28.3, 18.6.

tert-butyl ((2S,3R)-3-(benzyloxy)-1-(ethylamino)-1-oxobutan-2-yl) carbamate (12): obtained following the procedure A, 45.1 mg, 67% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.38–7.27 (m, 5H), 6.47 (br, 1H), 5.50 (d, *J*=6.2 Hz, 1H), 4.58 (q, *J*=11.4 Hz, 2H), 4.28–4.12 (m, 2H), 3.36–3.22 (m, 2H), 1.45 (s, 9H), 1.16 (d, *J*=6.3 Hz, 3H), 1.11 (t, *J*=7.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 168.3, 154.6, 136.8, 127.2, 126.5, 126.4, 78.8, 73.6, 70.3, 56.3, 33.1, 27.0, 14.2, 13.5. HRMS (ESI) calcd. for C₁₈H₂₈N₂O₄Na⁺: [M+Na]⁺: 359.1947, found: 359.1939.

N-(2-oxo-2-(phenethylamino) ethyl) benzamide (13)^[33]: obtained following the procedure A, 36.7 mg, 65% yield, white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.85–7.81 (m, 2H), 7.57–7.37 (m, 4H), 7.26–7.13 (m, 5H), 6.86 (brs, 1H), 4.08 (d, *J*=5.0 Hz, 2H), 3.54 (q, *J*=7.1 Hz, 2H), 2.82 (t, *J*=7.1 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 169.3, 168.0, 138.6, 133.3, 132.0, 128.7, 128.6, 127.2, 126.6, 43.9, 40.9, 35.6.

tert-butyl(1-((3-(methylthio)propyl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (14): obtained following the procedure B, 51.5 mg, 73% yield, yellow liquid. ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.13 (m, 5H), 6.04 (br s, 1H), 5.12 (br s, 1H), 4.28 (d, *J*=6.9 Hz, 1H), 3.26 (qt, *J*=13.3, 6.6 Hz, 2H), 3.03 (qd, *J*=13.5, 7.4 Hz, 2H), 2.52–2.31 (m, 2H), 2.03 (s, 3H), 1.66 (dd, *J*=13.1, 6.4 Hz, 2H), 1.40 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 171.2, 155.4, 136.8, 129.3, 128.7, 127.0, 80.2, 56.1, 38.7, 38.5, 31.4, 28.3,

28.3, 15.4. HRMS (ESI) calcd. for $C_{18}H_{28}N_2OSNa^+$: $[M + Na]^+$: 375.1718, found: 375.1707.

tert-butyl (2-(4-chlorophenyl) propyl) carbamate (15): obtained following the procedure A, 47.4 mg, 88% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.31–7.23 (m, 2H), 7.13 (d, $J = 8.3$ Hz, 2H), 4.48 (br s, 1H), 3.35 (dd, $J = 13.0, 6.4$ Hz, 1H), 3.15 (ddd, $J = 13.6, 8.2, 5.4$ Hz, 1H), 2.91 (dd, $J = 13.6, 6.7$ Hz, 1H), 1.41 (s, 9H), δ 1.23 (dd, $J = 7.0, 1.0$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 155.9, 142.7, 132.2, 128.7, 128.6, 79.2, 47.3, 39.6, 28.4, 19.0. HRMS (ESI) calcd. for $C_{14}H_{20}ClNO_2Na^+$: $[M + Na]^+$: 292.1080, found: 292.1073

N, N-bis (2-chloroethyl)-4-propylaniline (16): obtained following the procedure A, 46.8 mg, 90% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.08 (d, $J = 8.7$ Hz, 2H), 6.64 (d, $J = 8.7$ Hz, 2H), 3.76–3.57 (m, 8H), 2.53–2.46 (t, $J = 14.5$ Hz, 2H), 1.59 (dq, $J = 14.8, 7.4$ Hz, 2H), 0.93 (t, $J = 7.3$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 143.9, 132.3, 129.7, 112.3, 53.8, 40.5, 36.9, 24.8, 13.9. HRMS (ESI) calcd. for $C_{13}H_{20}Cl_2N^+$: $[M + H]^+$: 260.0973, found: 260.0963.

benzyl (S)-(2,4-dimethylpentyl)carbamate (17): obtained following the procedure A, 38.2 mg, 77% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.51–7.27 (m, 5H), 5.30–4.99 (m, 2H), 4.83 (s, 1H), 3.13 (dt, $J = 12.1, 5.9$ Hz, 1H), 2.96 (dt, $J = 13.4, 6.7$ Hz, 1H), 1.88–1.49 (m, 2H), 1.13 (ddd, $J = 13.8, 8.4, 5.7$ Hz, 1H), 1.07–0.95 (m, 1H), 0.98–0.71 (m, 9H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 156.6, 136.7, 128.5, 128.1, 128.1, 66.6, 47.4, 43.7, 31.2, 25.2, 23.4, 22.1, 17.6. HRMS (ESI) calcd. for $C_{15}H_{23}NO_2Na^+$: $[M + Na]^+$: 272.2626, found: 272.1616

benzyl ((1-methylcyclohexyl) methyl)carbamate (18): obtained following the procedure A, 42.7 mg, 82% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.41–7.27 (m, 5H), 5.12 (d, $J = 19.0$ Hz, 2H), 4.81 (br s, 1H), 3.02 (dd, $J = 18.0, 6.2$ Hz, 2H), 1.59–1.34 (m, 5H), 1.36–1.11 (m, 5H), 0.87 (s, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 156.81, 136.70, 128.54, 128.17, 128.12, 66.68, 51.34, 35.21, 34.29, 26.29, 23.00, 21.75. HRMS (ESI) calcd. for $C_{15}H_{24}NO_2^+$: $[M + H]^+$: 262.1807, found: 262.1802.

(Z)-heptadec-8-ene (19)^[9]: obtained following the procedure A, 27.2 mg, 67% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 5.44–5.28 (m, 2H), 2.06–1.99 (m, 4H), 1.37–1.24 (m, 22H), 0.89 (t, $J = 6.8$ Hz, 6H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 128.1, 30.1 (two singals), 28.0, 27.8, 27.6, 27.5 (two singals), 25.4, 20.9, 12.3.

(6Z, 9Z)-heptadeca-6, 9-diene (20)^[34]: obtained following the procedure A, 24.6 mg, 52% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 5.35–5.01 (m, 4H), 2.58 (t, $J = 6.4$ Hz, 2H), 1.86 (q, $J = 6.8$ Hz, 4H), 1.23–0.98 (m, 18H), 0.69 (m, $J = 6.9, 3.3$ Hz, 6H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 128.4, 126.2, 126.1, 30.1, 29.7, 27.9, 27.6, 27.5, 27.4, 25.5, 25.4, 23.8, 20.9, 20.8, 12.3, 12.3.

(E)-7-hydroxy-5-methoxy-4-methyl-6-(3-methylpent-2-en-1-yl) isobenzofuran-1 (3H)-one (21): obtained following the procedure A, 47.0 mg, 85% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.67 (s, 1H), 5.20 (s, 1H), 5.19–5.16 (m, 1H), 3.77 (s, 3H), 3.40 (d, $J = 6.9$ Hz, 2H), 2.15 (s, 3H), 1.99 (q, $J = 7.7$ Hz, 2H), 1.79 (s, 3H), 0.97 (t, $J = 7.5$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.0, 163.7, 153.7, 143.9, 137.7, 122.6, 120.4, 116.7, 106.4, 70.0, 61.0, 32.4, 22.6, 16.1, 12.6, 11.6. HRMS (ESI) calcd. for $C_{16}H_{20}O_4Na^+$: $[M + Na]^+$: 299.1259, found: 299.1250.

(1S/R, 4aS)-7-isopropyl-1, 4a-dimethyl-1, 2, 3, 4, 4a, 4b, 5, 6, 10, 10a-decahydrophenanthrene (22)^[35]: obtained following the procedure A, 36.2 mg, 80% yield, colorless liquid (d: r = 77: 23). 1H NMR (400 MHz, $CDCl_3$) δ 5.78 (s, 1H, vinyl H), 5.52–5.37 (m, 1H, vinyl H), 2.30–2.03 (m, 4H), 1.82 (tq, $J = 10.6, 3.6$ Hz, 5H), 1.66–1.39 (m, 6H), 1.06–0.96 (m, 10H), 0.78 (s, 2.3H, major diastereoisomer), 0.72 (s, 0.7H, minor diastereoisomer).

1, 4-dimethyl-2-((4-methylpentyl) oxy) benzene (23): obtained following the procedure A, 34.3 mg, 83% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.00 (d, $J = 7.4$ Hz, 1H), 6.65 (d, $J = 7.6$ Hz, 1H), 6.62 (s, 1H), 3.92 (t, $J = 6.5$ Hz, 2H), 2.30 (s, 3H), 2.18 (s, 3H), 1.84–1.75 (m, 2H), 1.62 (dp, $J = 13.3, 6.7$ Hz, 1H), 1.39–1.32 (m, 2H), 0.92 (d, $J = 6.6$ Hz, 6H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 157.1, 136.4, 130.3, 123.6, 120.5, 112.0, 68.1, 35.3, 27.8, 27.3, 22.6, 21.4, 15.8. HRMS (ESI) calcd. for $C_{14}H_{23}O^+$: $[M + H]^+$: 207.1749, found: 207.1739.

4-propoxybenzaldehyde (24)^[36]: obtained following the procedure B, 32.0 mg, 73% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 9.88 (s, 1H), 8.37–7.54 (m, 2H), 7.15–6.80 (m, 2H), 4.01 (t, $J = 6.6$ Hz, 2H), 2.11–1.77 (m, 2H), 1.06 (t, $J = 7.4$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 190.9, 164.3, 132.0, 129.7, 114.8, 69.9, 22.4, 10.5.

2-(4-ethoxyphenyl)-4, 4, 5, 5-tetramethyl-1, 3, 2-dioxaborolane (25)^[37]: obtained following the procedure B, 36.2 mg, 90% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.74 (d, $J = 8.6$ Hz, 2H), 6.88 (d, $J = 8.7$ Hz, 2H), 4.05 (q, $J = 7.0$ Hz, 2H), 1.41 (t, $J = 7.0$ Hz, 3H), 1.33 (s, 12H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 161.5, 136.5, 113.8, 83.5, 63.2, 24.9, 14.8.

4, 4, 5, 5-tetramethyl-2-(4-(pentyloxy)phenyl)-1, 3, 2-dioxaborolane (26): obtained following the procedure B, 41.6 mg, 72% yield, white solid. 1H NMR (400 MHz, $CDCl_3$) δ 7.77 (d, $J = 8.5$ Hz, 2H), 6.92 (d, $J = 8.5$ Hz, 2H), 4.00 (t, $J = 6.6$ Hz, 2H), 2.01–1.69 (m, 2H), 1.61–1.10 (m, 16H), 0.96 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 162.34, 137.08, 136.22, 114.45, 84.11, 68.36, 29.51, 28.78, 25.45, 23.06, 14.63. 1HRMS (ESI) calcd. for $C_{17}H_{27}BO_3^+$: $[M + H]^+$: 291.2132, found: 291.2117.

3-methyl-1-tosylpyrrolidine (28)^[38]: obtained following

the procedure A, 33.0 mg, 69% yield, white solid. ^1H NMR (400 MHz, CDCl_3) δ 7.69 (d, $J=8.2$ Hz, 2H), 7.30 (d, $J=8.0$ Hz, 2H), 3.40 (dd, $J=9.7, 7.2$ Hz, 1H), 3.36–3.29 (m, 1H), 3.23–3.16 (m, 1H), 2.73 (dd, $J=9.7, 7.8$ Hz, 1H), 2.42 (s, 3H), 2.10 (td, $J=14.7, 7.1$ Hz, 1H), 1.93–1.84 (m, 1H), 1.33 (dq, $J=12.3, 8.3$ Hz, 1H), 0.90 (d, $J=6.7$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 143.25, 134.02, 129.60, 127.53, 54.77, 47.62, 33.32, 33.26, 21.55, 17.65.

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Conflict of interest

The authors declare no conflict of interest.

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无需自由基引发剂参与实现可见光诱导 Barton 脱羧

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摘要: 报道了一种可见光诱导 Barton 脱羧的新方法, 在不需外加自由基引发剂的条件下, 可以实现一系列一级、二级、三级以及氨基酸、多肽类氧化还原性羧酸酯脱羧, 而且许多含有羧基的天然产物及药物分子也可以很好地发生转化, 反应条件温和、底物范围广, 具有良好的收率和官能团兼容性. 初步机理实验研究表明, 三(三甲基硅基)硅烷在该反应体系中不仅作为氢源, 同时也作为电子给体和氧化还原性羧酸作用形成电子供体-受体复合物, 反应经历自由基历程.

关键词: 可见光诱导; 电子供体-受体复合物; 脱羧; 自由基; 三(三甲基硅基)硅烷